



Quelques perspectives dans l'étude expérimentale du transport colloidal en milieu poreux complexe

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Colloidal transport in porous materials

- **Filtration (transport, glogging)**
- **Chromatography**
- **Micro and nano fluidic**
- **Aquifer**
- **Porous rocks (sandstone,dolomite,.....)**
- **Soil decontamination by colloids injection**

Direct observation of colloidal transport: imaging colloidal trajectories

Not a new idea! (1907)

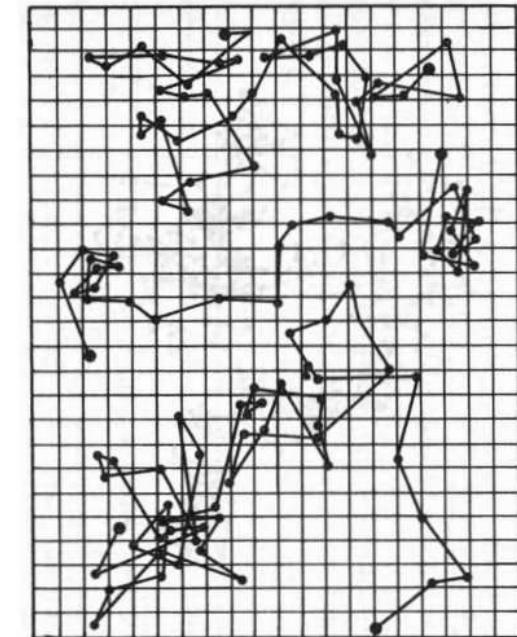


JEAN PERRIN
1870-1942

Jean Perrin



Microphotographie de la répartition en hauteur de sphères de résines en suspension dans l'eau.
Photographie reproduisant un cliché de Perrin, réalisé par le palais de la Découverte pour la présentation
de ses expériences sur le mouvement brownien.



Dessin représentant le mouvement brownien de trois particules distinctes. Au moyen d'un projecteur, l'image de la solution est projetée sur un écran dépoli et quadrillé. À intervalles réguliers, les positions successives d'une particule sont notées par des points, ce qui permet de calculer le déplacement moyen des particules. Jean Perrin, *Mouvement brownien et réalité moléculaire*, in *Annales de chimie et de physique*, série 8, vol. 18, 1909, p. 81.

$$D_{self} = \frac{k_B T}{6\pi\eta R}$$

Self-Limited Accumulation of Colloids in Porous Media

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As a model porous medium we use monodisperse borosilicate beads (negatively charged, $63 \mu\text{m}$ average diameter) randomly packed (porosity $\phi_0 = 0.38$) in a square glass capillary (width $w = 1.0 \text{ mm}$), and we invade this medium with positively charged, fluorescent, monodisperse (diameter $d_p = 1.0 \mu\text{m}$) latex colloids. The suspensions are made at constant dilute concentration $C_0 = 20 \text{ mg mL}^{-1}$ in a mix of deionized water and DMSO to match the refractive index of the beads [28]. This allows us to visualize the particles inside the porous medium by

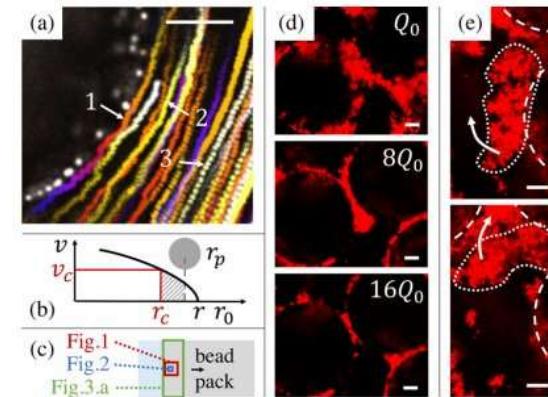


FIG. 2. Local transport and adsorption mechanisms. All scale bars are $10 \mu\text{m}$. (a) Time projection of confocal imaging, with the suspension flowing upwards. $I = 10^{-6} \text{ M}$. Steady particles appear as white dots, while moving particles are represented by their trajectories. Selected trajectories: particle 1 along wall, 2 close to wall, 3 far from wall. (b) Schematic velocity profile in a pore of initial and current radii r_0 and r . Dashed area shows positions favorable to adsorption: $[r_c : r - r_p]$. (c) Schematics of the three scales used in Figs. 1, 2 and 3(a). (d) Selected area under steady state for different flow rates [$Q_0; 8Q_0; 16Q_0$], at $I = 10^{-1} \text{ M}$. (e) Cluster formed on a surface at $I = 10^{-1} \text{ M}$ (top), detaching and moving to a more stable position (bottom). Dashed lines highlight bead surfaces.

**Very nice experiment but:
Need for relative light transparency of the pore network
Need to observe a large number of trajectories in order to perform
in a post processing analysis, an ensemble average.**

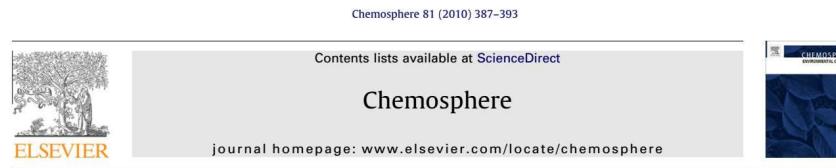
Some needs in case of completely opaque and/or multidiffusif materials

- No direct visualization of colloidal trajectories.
- Need for a temporal resolution.
- Need to perform an experimental ensemble average.
- Need to extract meaningful “descriptors” either at macroscopic or at mesoscopic length scales

In this talk:

- Macroscopic experiments: Breakthrough curves
- Use of the Magnetic resonance imaging
- X-ray photon correlation spectroscopy (XPCS)
- Part 4 not available for actual publication

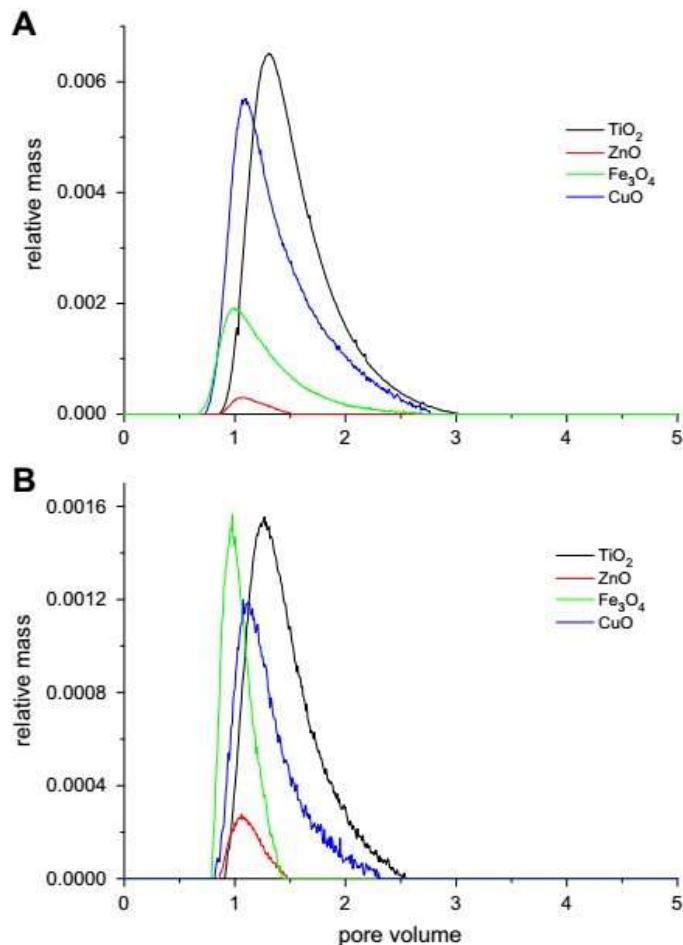
Part 1: Macroscopic experiments: Breakthrough curves



Transport of metal oxide nanoparticles in saturated porous media

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$$\frac{\partial}{\partial t} C = D \frac{\partial^2 C}{\partial x^2} - v_p \frac{\partial C}{\partial x} - kC$$

Fig. 4. Breakthrough curves for metal oxide nanoparticles at pH 7 with (A) 0.01 M NaCl and (B) 0.1 M NaCl.

Part 2: A power tool – the Magnetic Resonance Imaging

Int. J. Environ. Sci. Technol. (2015) 12:3373–3384
DOI 10.1007/s13762-015-0767-4



ORIGINAL PAPER

Characterization of nanoparticle transport through quartz and dolomite gravels by magnetic resonance imaging

S. Lakshmanan · W. M. Holmes · W. T. Sloan ·
V. R. Phoenix

$$\frac{S}{S_0} = \exp\left(\frac{-t_e}{T_{2,i}}\right)$$

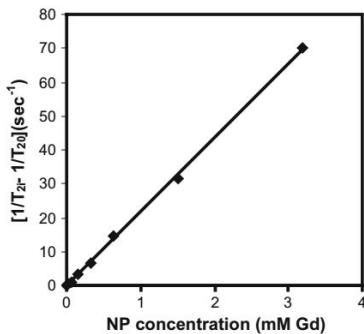


Fig. 1 Variation in the change of transverse relaxation rate with respect to *Gd* NP concentration (NP concentration shown as mM of *Gd*)

$$[C] = \frac{1}{R} \left[\frac{1}{T_{2,i}} - \frac{1}{T_{2,0}} \right]$$

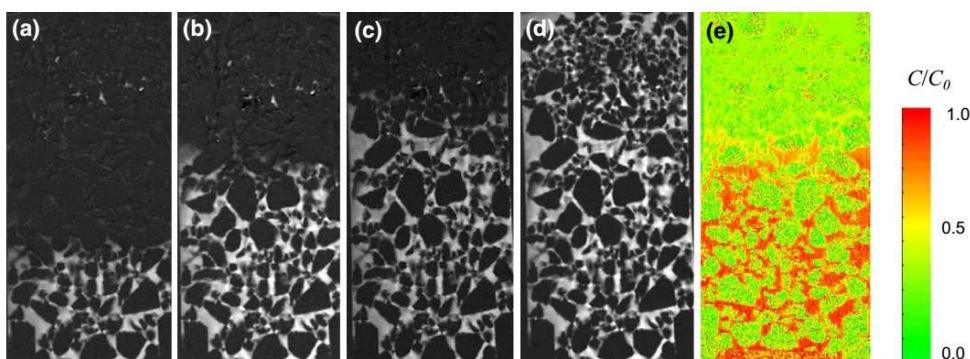
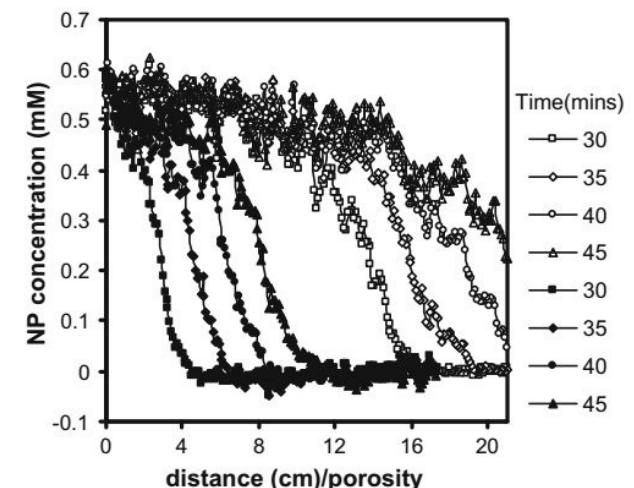


Fig. 3 T_2 -weighted images of *Gd* NP in dolomite gravel at 35, 45, 65 and 75 min denoted by (a), (b), (c) and (d), respectively; NP transport is shown by bright regions. Figure 3e shows calibrated image of *Gd* NP concentration at 55 min



Comparison exp-model

$$\frac{\partial}{\partial t} C = D \frac{\partial^2 C}{\partial x^2} - v_p \frac{\partial C}{\partial x} - kC \quad (4)$$

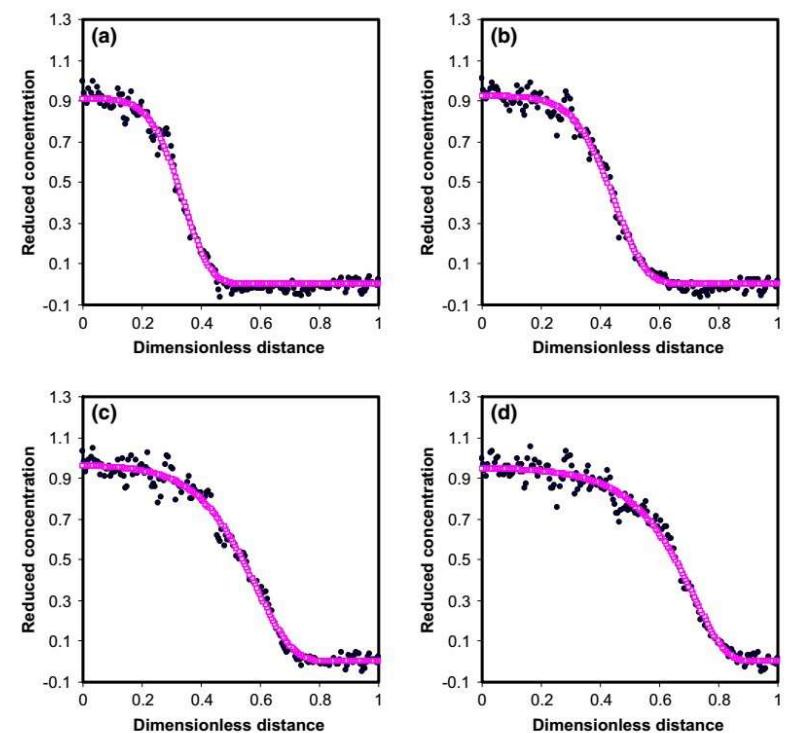
where C is the concentration, t is time, x is distance from the inlet, v_p average particle velocity, D dispersion coefficient and k is the decay coefficient. We assume a semi-infinite domain with constant concentration C_0 at the inlet.

$$C(x, t) = C_0 \text{ when } x = 0, t \geq 0 \quad (5a)$$

$$C(x, t) = C_0 \text{ when } 0 < x < \infty, t = 0 \quad (5b)$$

$$\frac{dC(x, t)}{dx} = 0 \text{ when } x = \infty \quad (5c)$$

Fig. 7 Gd NP concentration profile of dolomite gravel experiment at **a** 40 min, **b** 45 min, **c** 50 min, **d** 55 min. Experimental data (closed symbol) and CXTFIT data (open symbol)



See also Lehoux, S. Rodts et al, Phys. Rev E, 94, 053107(20216)

Using X rays beams to probe colloidal dynamics

Time correlation of 2D images

- 2D scattering images (XPCS)

Part 3 : XPCS _ X ray photon Correlation Spectroscopy

**“Extension” in the X-rays domain of the DLS (homodyne detection):
Time resolved 2D scattering technique**

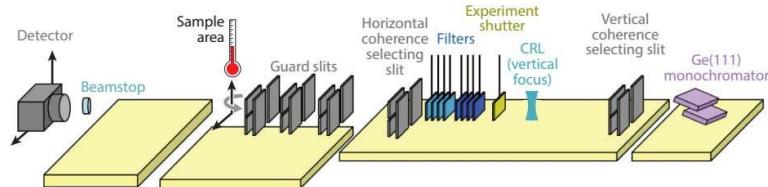
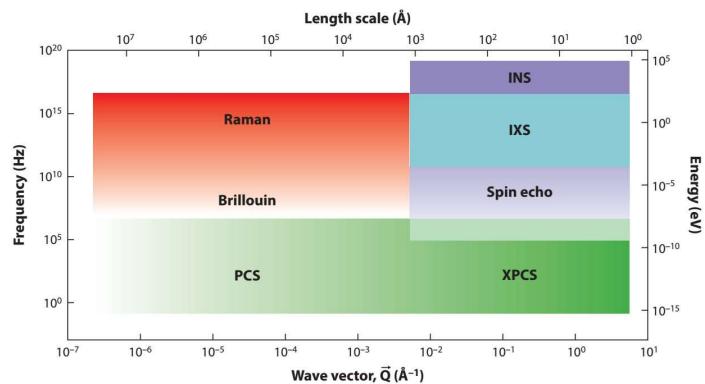


Figure 4

Schematic of the small-angle XPCS setup in enclosure 8-ID-I at the Advanced Photon Source. From right to left, 8-ID-I contains a Ge(111) monochromator, transverse coherence length-selecting slits, a compound refractive lens (CRL) for 1D vertical focusing at the sample position, a sample platform capable of supporting a wide variety of sample environments, a 4-m-long exit flight path, and a variety of area detectors. The monochromator is 65 m from the source, and the sample area is 68 m from the source. The detector and beamstop are 4 m downstream of the sample area.

No multidiffusive scattering. Large q range. Use of the Sieger theorem

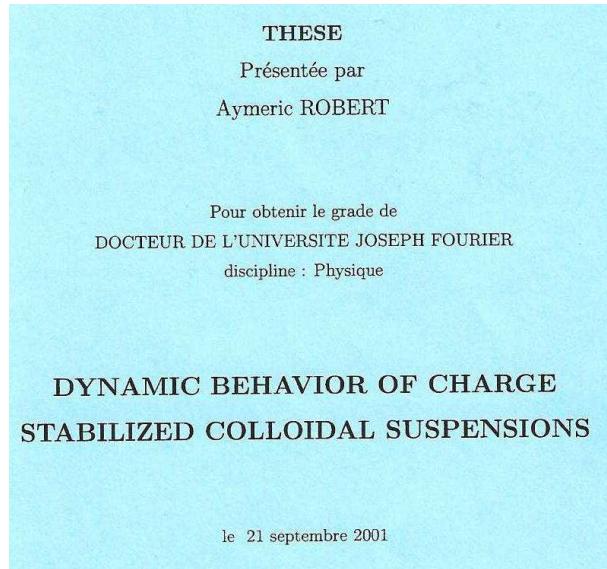
$$g_2(Q, \tau) = \frac{\langle I(Q, t)I(Q, t + \tau) \rangle}{\langle I(Q) \rangle^2} = \beta(Q)g_1^2(Q, \tau) + 1,$$

Intermediate scattering function (ISF)

$$g_1(\vec{q}, \tau) = \left\langle \frac{1}{N} \sum_{i=1}^N \sum_{j=1}^N \exp(i\vec{q}(\vec{r}_j(t_0 + \tau) - \vec{r}_i(t_0))) \right\rangle_{t_0}$$

One the first application:

Probing the dynamics of very concentrated colloidal suspensions:



THE JOURNAL OF CHEMICAL PHYSICS **137**, 114504 (2012)

Structure and short-time dynamics in concentrated suspensions of charged colloids

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$$D_m(Q) = - \lim_{\tau \rightarrow 0} \frac{1}{Q^2} \frac{\partial}{\partial \tau} \ln g_1(Q, \tau) = D_0 \frac{H_m(Q)}{S_m(Q)},$$

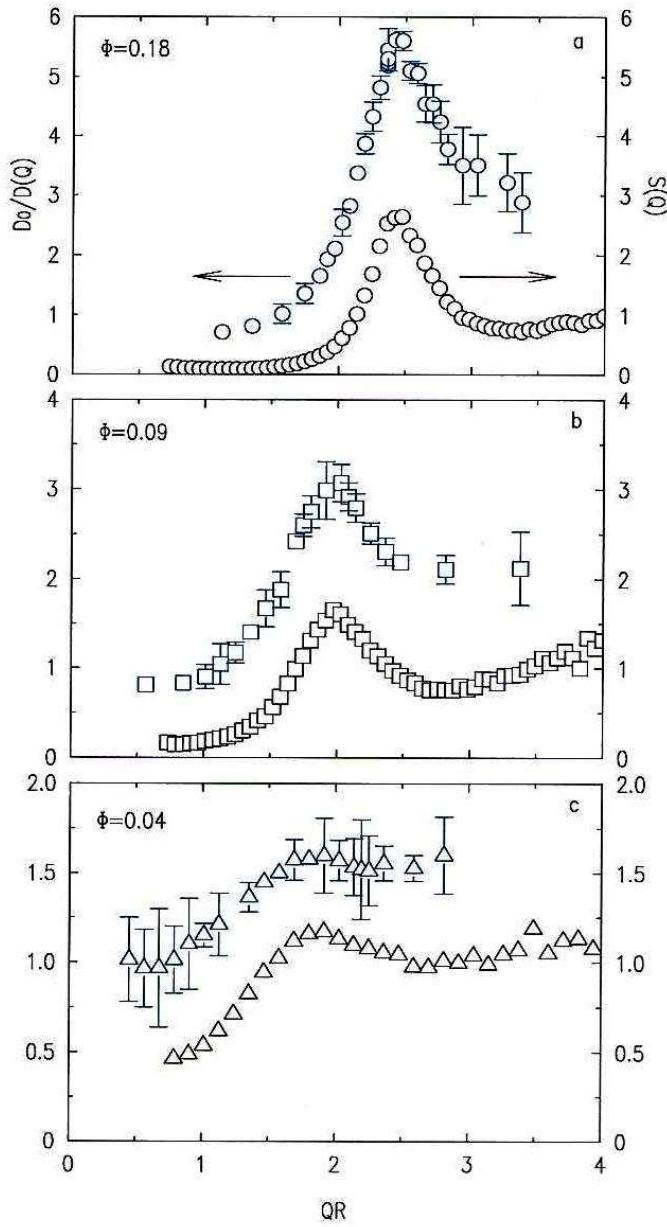


Figure 4.15: Static structure factor (\circ) for the colloidal suspensions of volume fraction $\phi = 0.18$ (a), 0.09 (b) and 0.04 (a). The blue symbols shows the Q -dependence of the normalized inverse effective diffusion coefficient $D_0/D(Q)$.

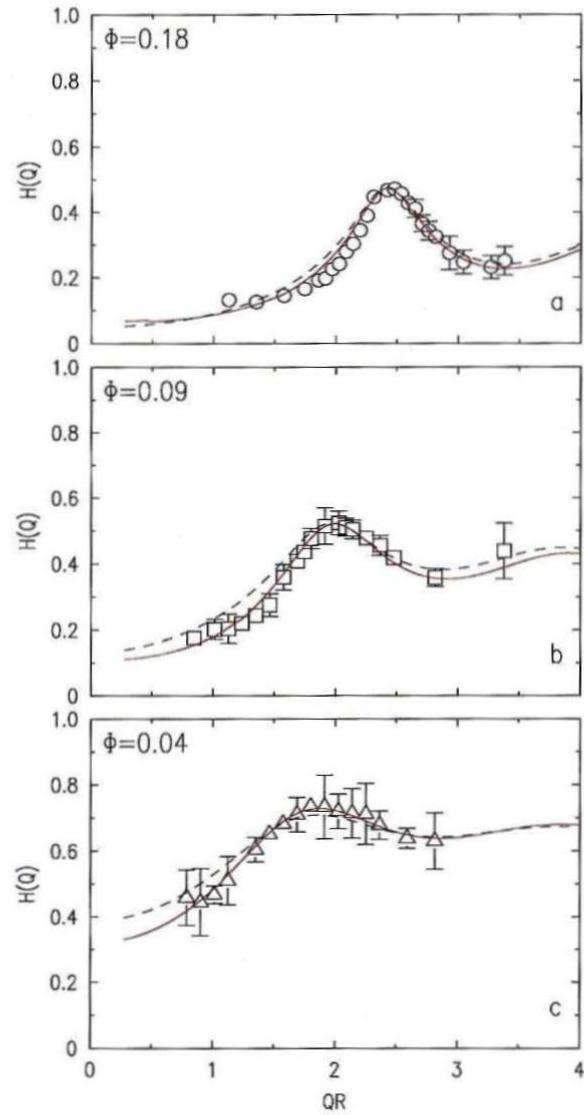


Figure 4.20: Hydrodynamic functions compared to two model calculations. The dashed lines are the result of the fit of eq. 4.3.8 (small Q approximation) to the data using the measured static structure factor as an input and with \tilde{D} being the only fit parameter. The solid lines are the results of the fit of eq. 4.3.2 ($\delta\gamma$ -expansion) using the measured static structure factor as an input and with α being the scaling parameter (see text).

Extremely Slow Diffusion of Gold Nanoparticles under Confinement in Mesoporous Silica

Ramona Mhanna, Michael Giroux, Kenneth J. T. Livi, Chao Wang, Andrei Fluerasu, Lutz Wiegart, Yugang Zhang, Mark Sutton, and Robert L. Leheny*



Cite This: *J. Phys. Chem. C* 2022, 126, 3614–3622



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$$g_2(q, t) = 1 + \beta(X^2|g_{1,\text{Au}}(q, t)|^2 + 2X(1 - X)\text{Re}[g_{1,\text{Au}}(q, t)g_{1,\text{SBA}}^*(q, t)] + (1 - X)^2|g_{1,\text{SBA}}(q, t)|^2)$$

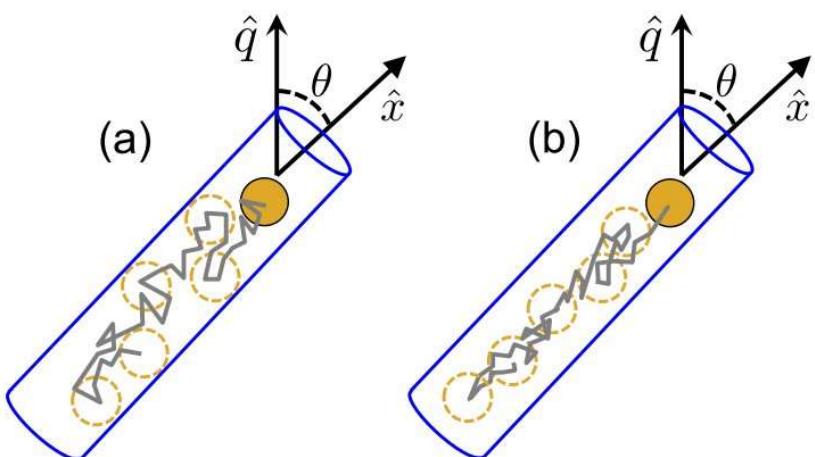
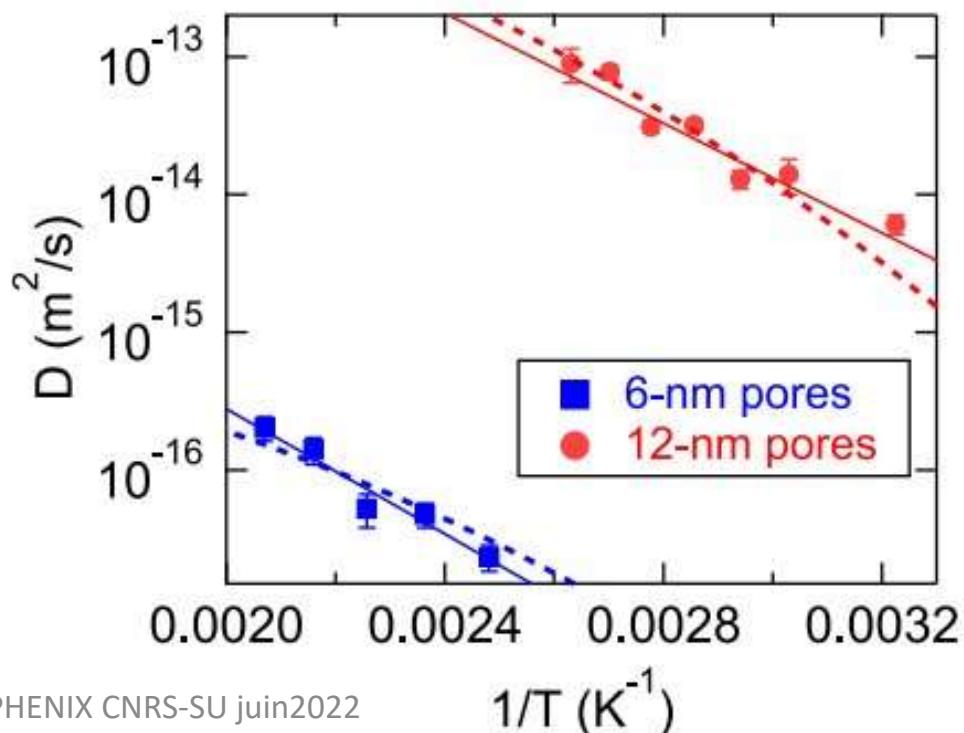


Figure 3. Schematics depicting different scenarios for a nanoparticle diffusing in glycerol within an SBA-15 pore. (a) Particle's trajectory is punctuated by repeated adsorption to the pore walls. (b) Particle is restricted to the central region of the pore because of an immobile layer of glycerol adjacent to the walls.



To sum-up for XPCS

- * Time correlation of 2D scattering images.
- * Origin of 2D image contrast: Atomic X-ray scattering section
- * Need to have a highly coherent X ray (lateral and longitudinal)
(to get β close to 1)
- * Can be performed on synchrotron beam lines (ID2, ID10 for ESRF)
- * Need to check for radiation damages

Part 4

Not actually available